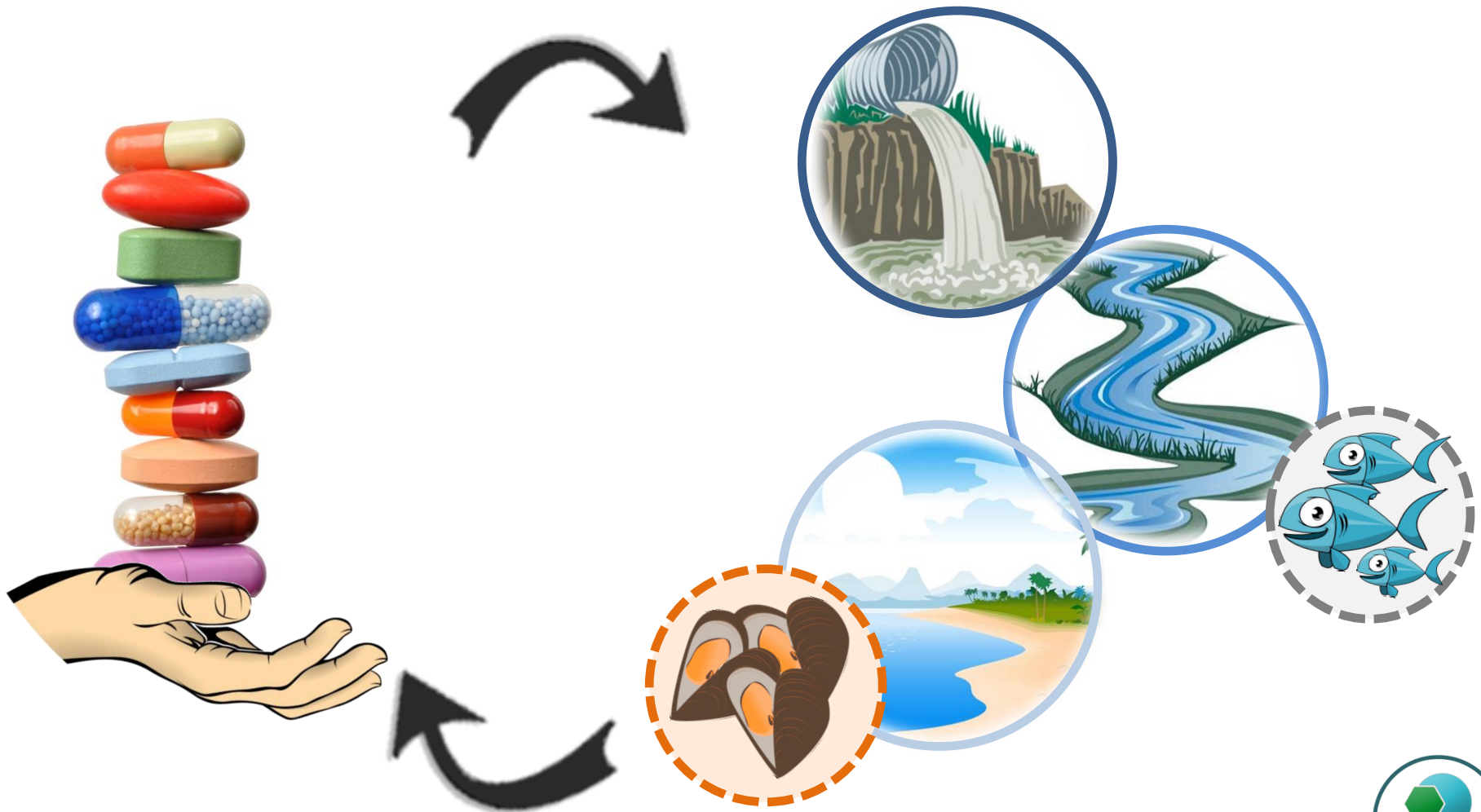


# Determination of fluoroquinolones in fish tissues, biological fluids and environmental waters

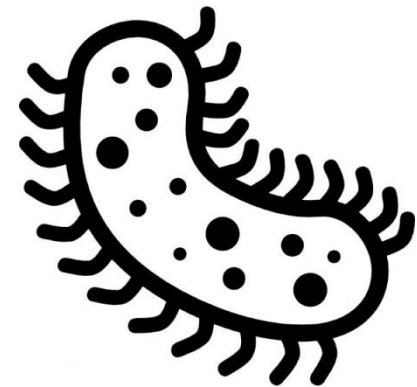
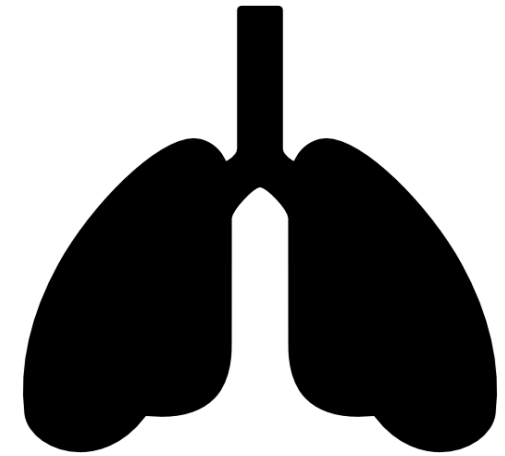
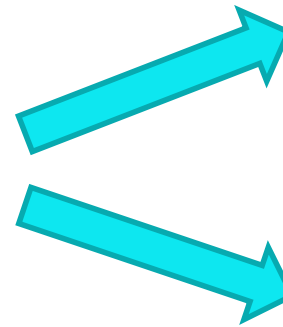
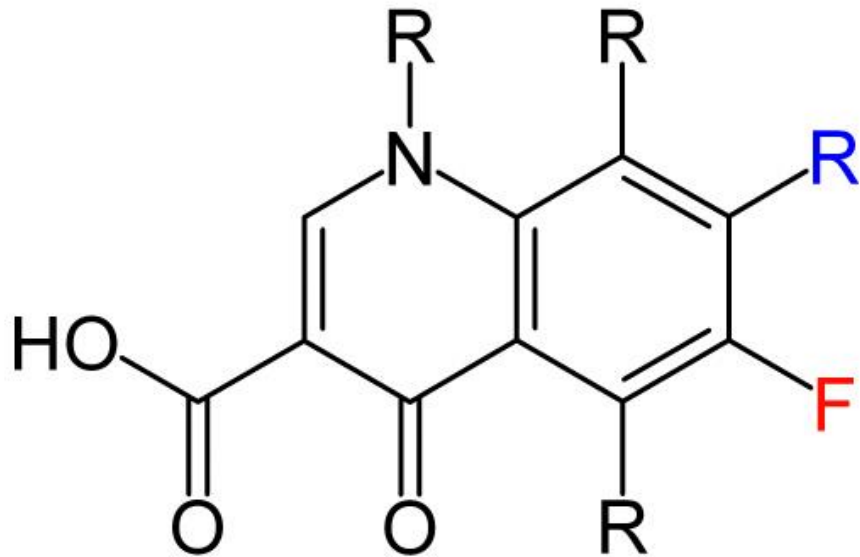
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# Pharmaceuticals



# Fluoroquinolones (FQs)



# Fluoroquinolones: effects



# Fluoroquinolones: method development

Lack of analytical methods



# Objectives



1

Optimization LC-MS/MS analysis of FQs

2

Optimization of the extraction: fish tissue/biofluids + environmental water

3

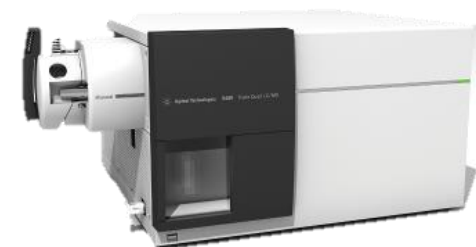
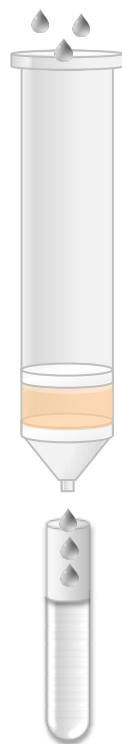
Application to real samples

# Experimental part

## Solid-Phase Extraction (SPE)

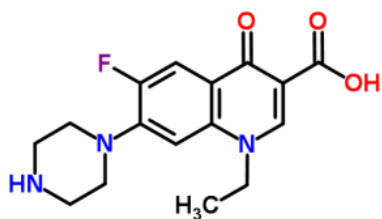


**Focused Ultrasound  
Solid-Liquid Extraction  
(FUSLE)**

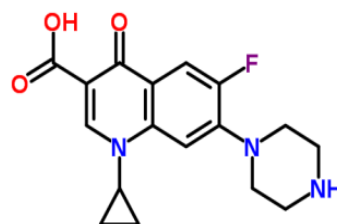


**Liquid Chromatography  
tandem Mass Spectrometry  
(LC-MS/MS)**

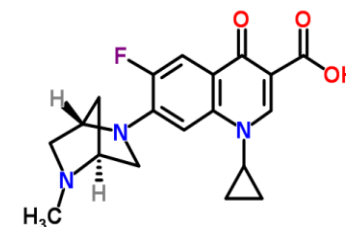
# Fluoroquinolones



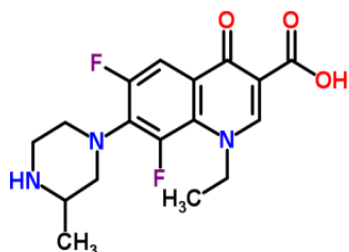
Norfloxacin (NORF)



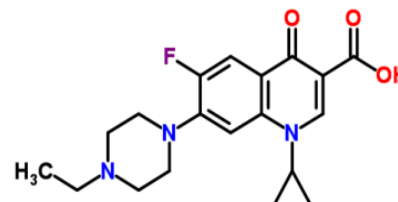
Enofloxacin (ENO)



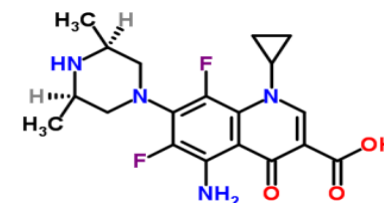
Pefloxacin (PEFLO)



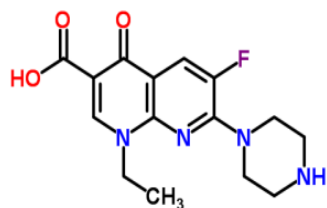
Ofloxacin (OFLO) and Levofloxacin (LEVO)



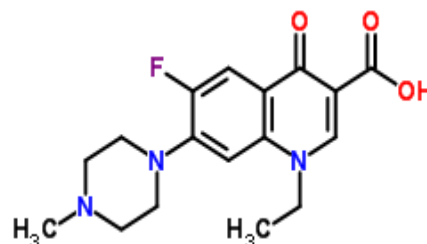
Ciprofloxacin (CIPRO)



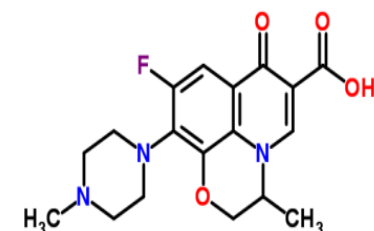
Danofloxacin (DANO)



Lomefloxacin (LOME)



Enrofloxacin (ENRO)



Sparfloxacin (SPAR)

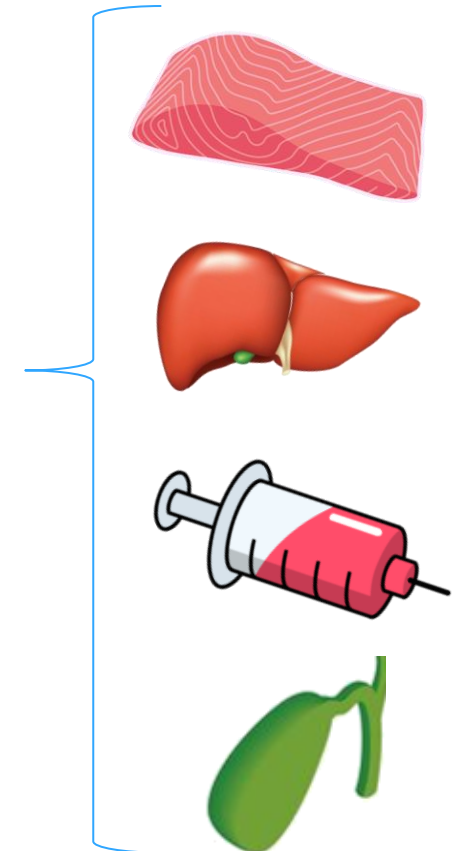
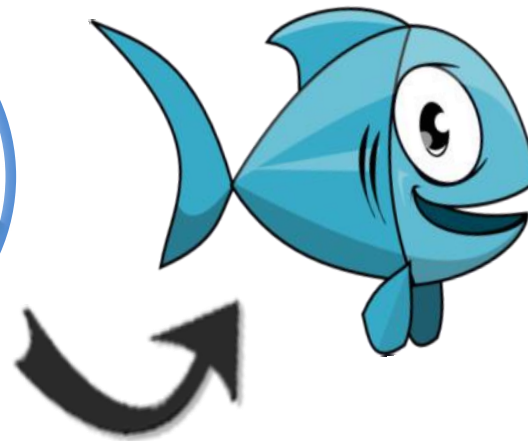


# Environmental samples

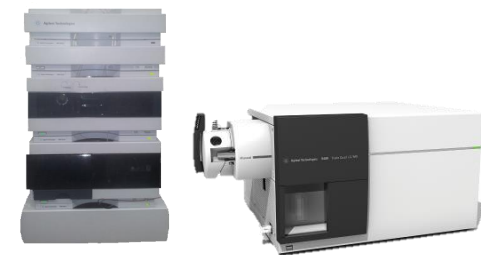
## Environmental waters



## Fish tissues and biofluids



# Results and Discussion



1

Optimization LC-MS/MS analysis of FQs

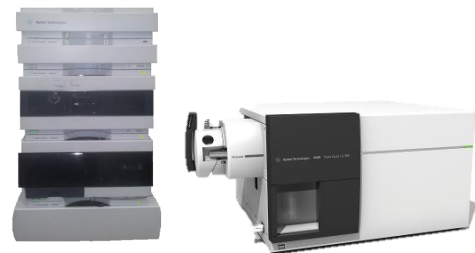
2

Optimization of the extraction: fish tissue/biofluids + environmental water

3

Application to real samples

# Optimization of LC-MS/MS



- Column temperature: 35 °C
- Flow rate: 0.3 mL/min
- Gradient
- Max. injection volume: 3 μL

Chromatographic parameters

- Capillary voltage: 3000 V
- Nebulizer pressure: 30 psi
- Drying gas flow: 8 L/h
- Drying gas temp.: 300 °C

Electrospray ionization parameters

- Fragmentor and collision energy

MS/MS parameters

# Fragmentor and collision energy

Analyte	SRM transitions
NORF	320 (104 V) → 302/231/282 (17/41/29 eV)
ENO	321 (104 V) → 303/232/204 (17/37/45 eV)
PEFLO	334 (104 V) → 316/290/233 (17/13/25 eV)
OFLO/LEVO	362 (104 V) → 318/261/344 (17/25/17 eV)
CIPRO	332 (104 V) → 314/231/288 (17/41/13 eV)
DANO	358 (104 V) → 340/82/255 (21/45/41 eV)
LOME	352 (104 V) → 265/308/334 (21/13/17 eV)
ENRO	360 (104 V) → 342/316/286 (17/17/37 eV)
SPAR	393 (104 V) → 349/292/375 (17/21/17 eV)

# Results and Discussion

1

Optimization LC-MS/MS analysis of FQs

2

Optimization of the extraction: fish tissue/biofluids + environmental waters

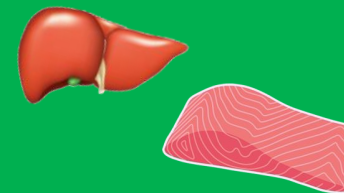
3

Application to real samples

# Optimization of the extraction

2.1

Fish tissues: liver and muscle



2.2

Fish biofluids: plasma and bile



2.3

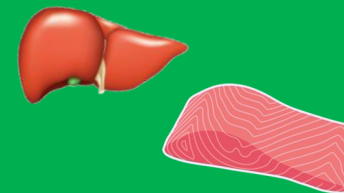
Environmental waters: seawater, estuarine water and WWTP water



# Optimization of the extraction

2.1

Fish tissues: liver and muscle



2.2

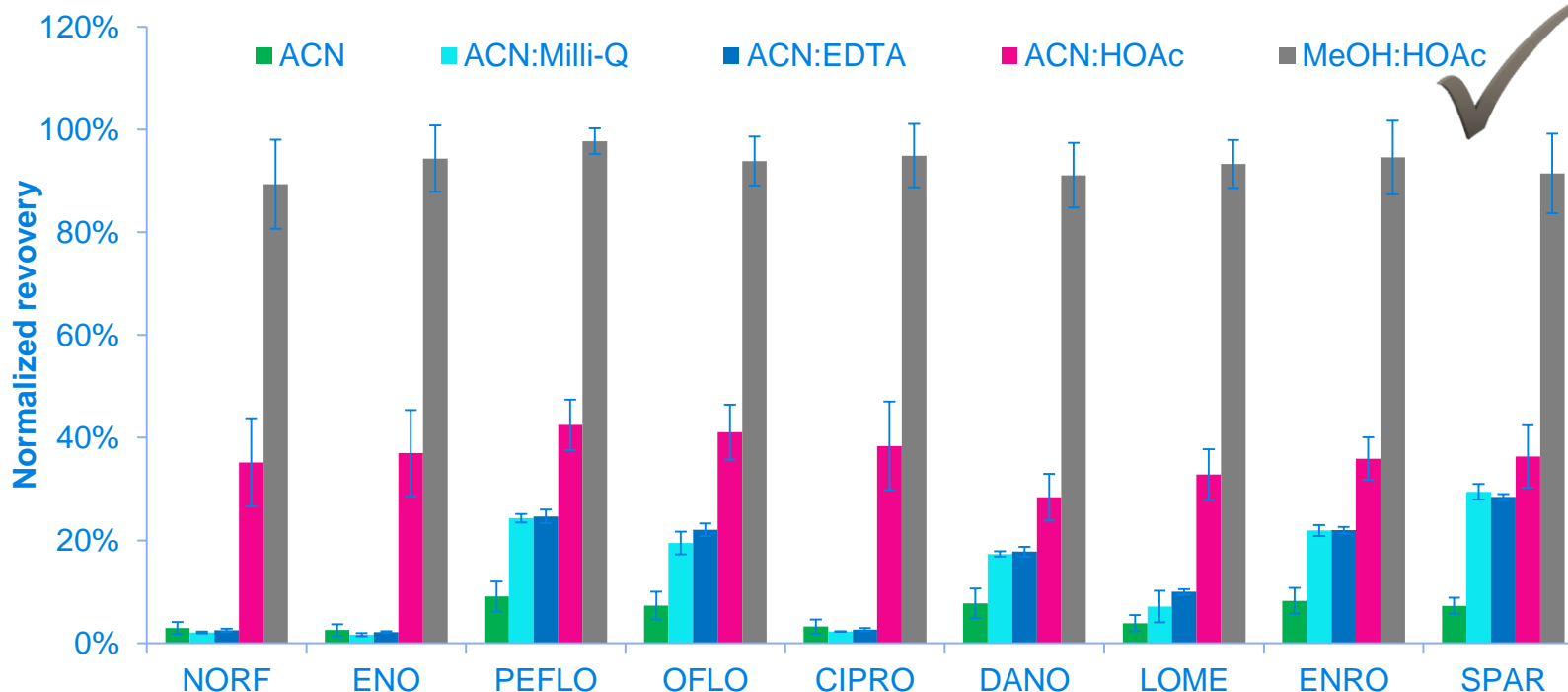
Fish biofluids: plasma and bile

2.3

Environmental waters: seawater, estuarine water and WWTP water

# Fish tissues: FUSLE

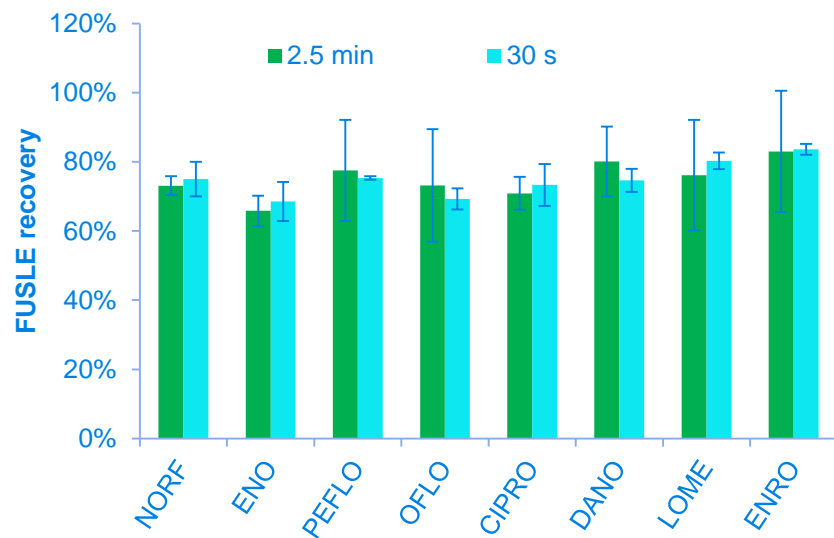
- Extraction solvent





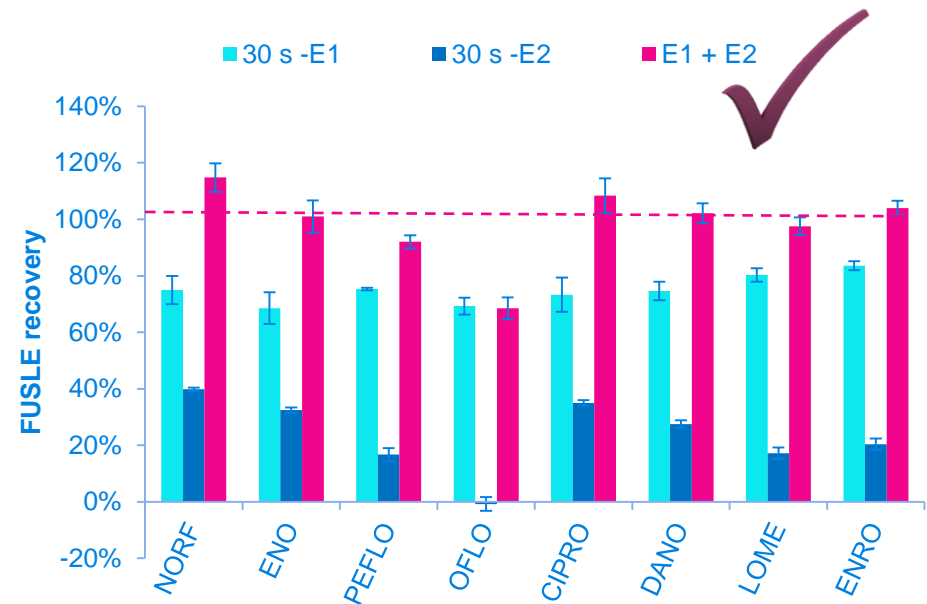
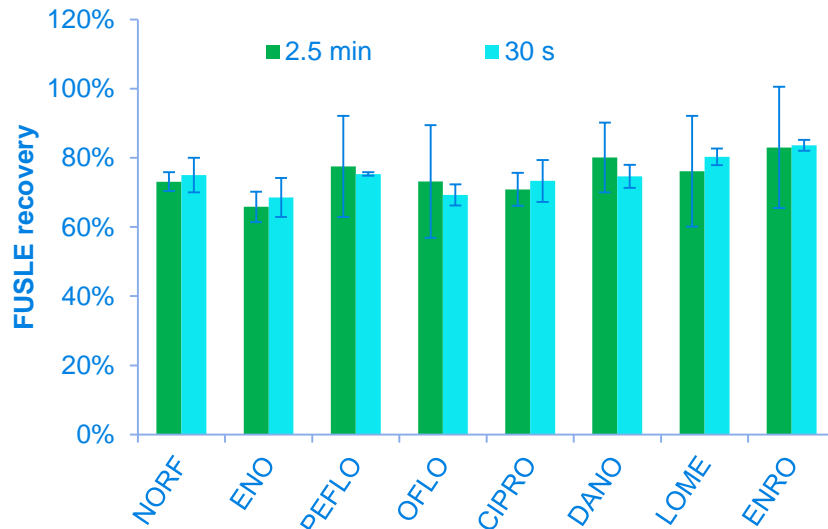
# Fish tissues: FUSLE

- Extraction time



# Fish tissues: FUSLE

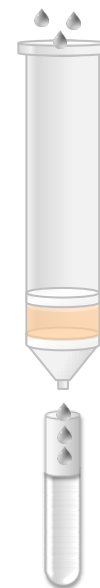
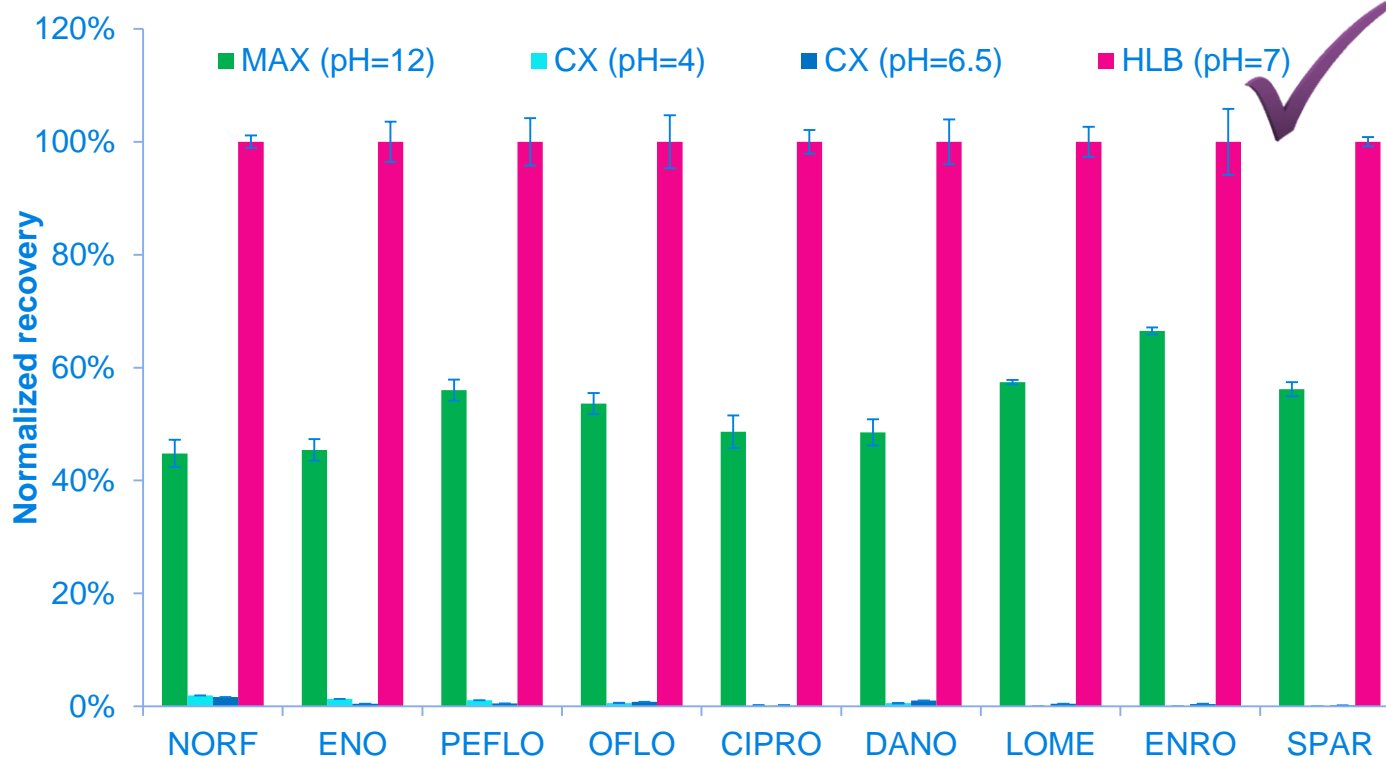
- Extraction time



Optimum: 30 s x 2

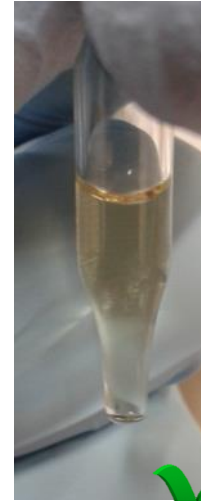
# Solid-Phase Extraction (SPE)

- Sorbent phase



# Liver: dirty extracts

Muscle

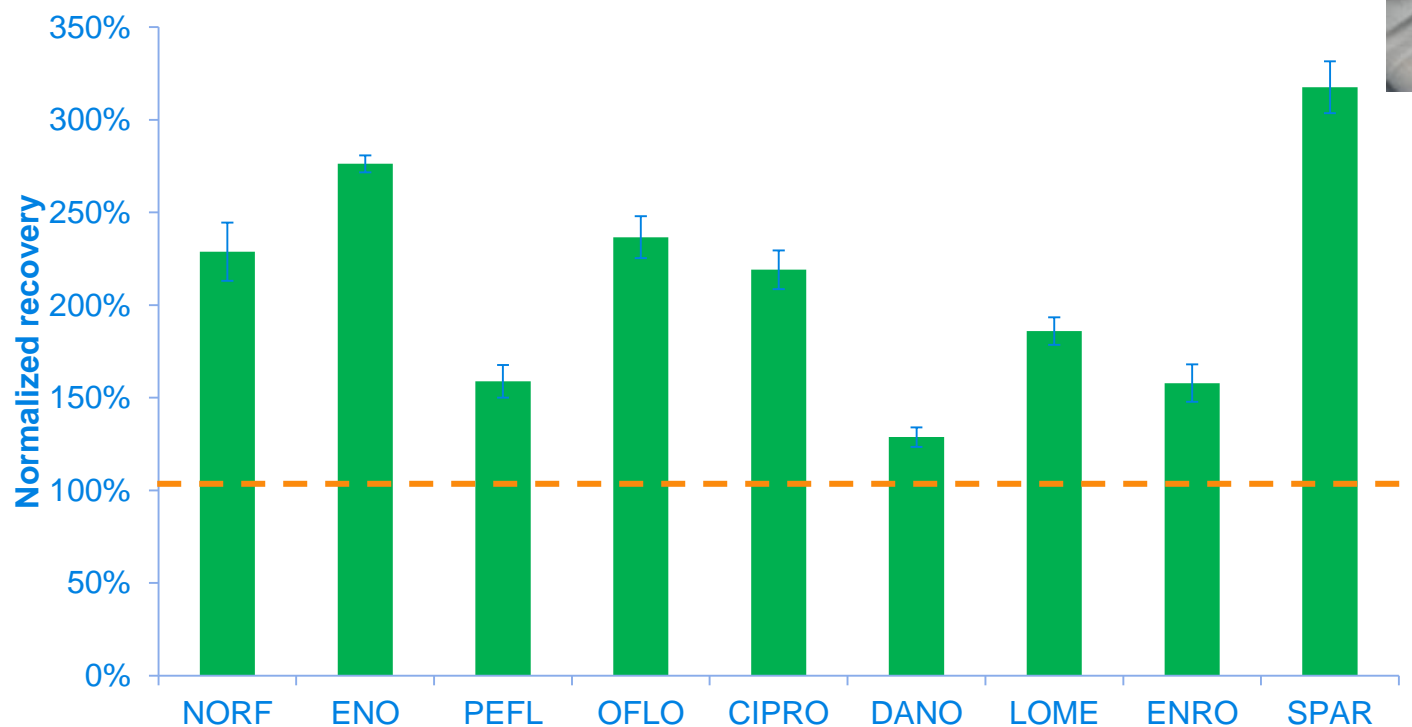


Liver



# Liver: dirty extracts

- Positive matrix effect



Liver



# Liver: extra step to remove lipids

- Clean-up strategies

# Liver: extra step to remove lipids

- Clean-up strategies

1. Increase HLB phase



# Liver: extra step to remove lipids

- Clean-up strategies

1. Increase HLB phase



2. Florisil-HLB





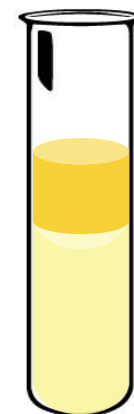
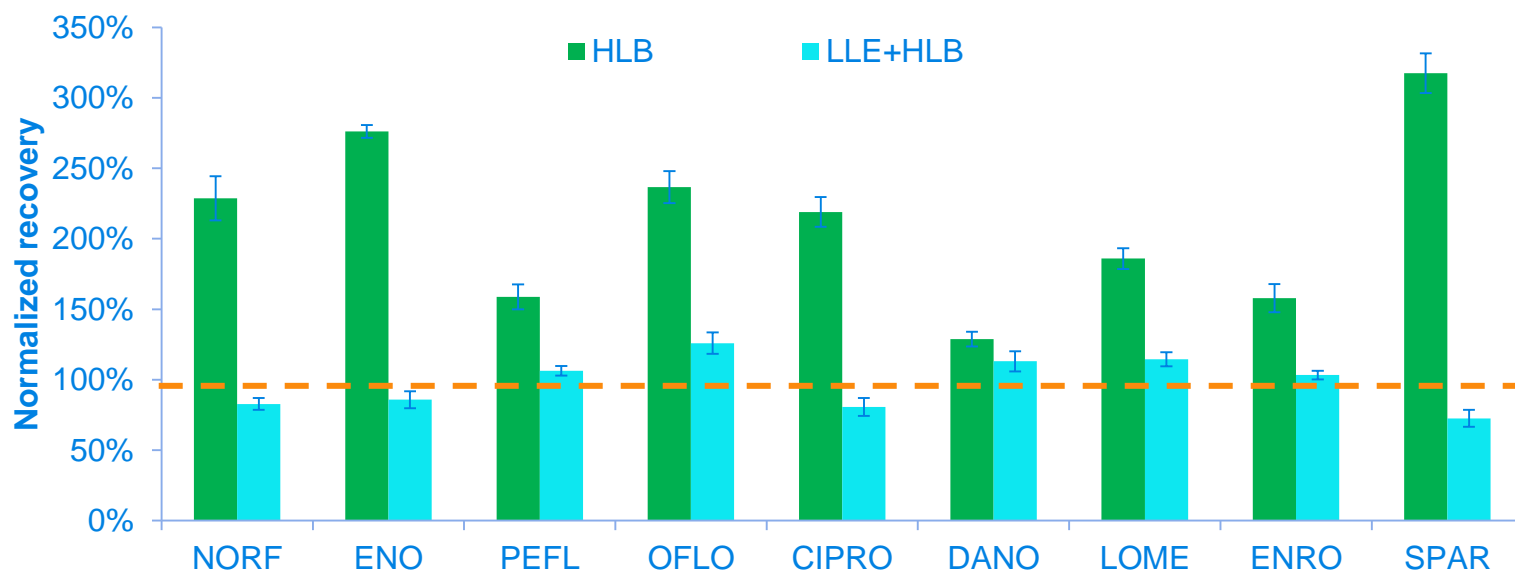
# Liver: extra step to remove lipids

## • Clean-up strategies

1. Increase HLB phase 

2. Florisil-HLB 

3. Liquid-liquid extraction (LLE) prior to HLB 



# Optimization of the extraction

2.1

Fish tissues: liver and muscle

2.2

Fish biofluids: plasma and bile



2.3

Environmental waters: seawater, estuarine water and WWTP water

# Biofluids: SPE

- HLB: signal suppression **X**



# Biofluids: SPE

- HLB: signal suppression
- MIPs cartridges



# Optimization of the extraction

2.1

Fish tissues: liver and muscle

2.2

Fish biofluids: plasma and bile

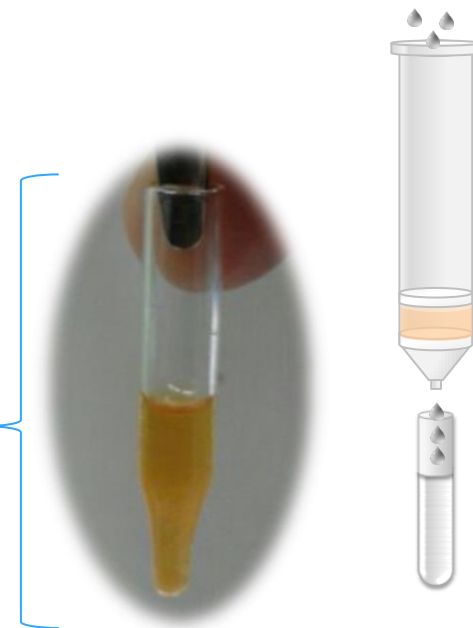
2.3

Environmental waters: seawater, estuarine water and WWTP water



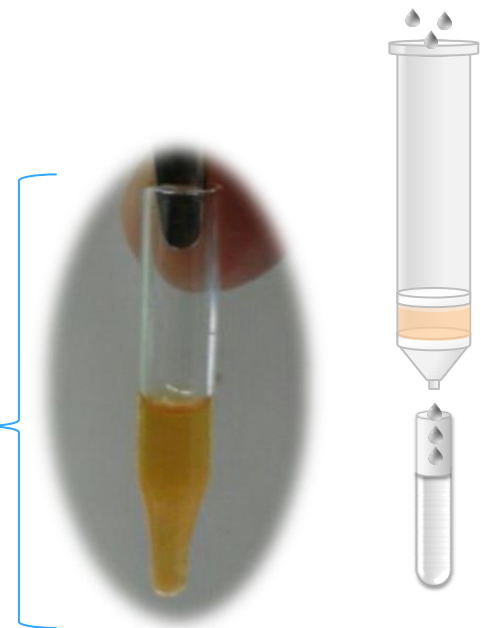
# RP-SPE (Oasis HLB)

- Seawater ✓
- Estuarine and effluent: dirty extracts
  - Signal suppression ( > 50 %)



# RP-SPE (Oasis HLB)

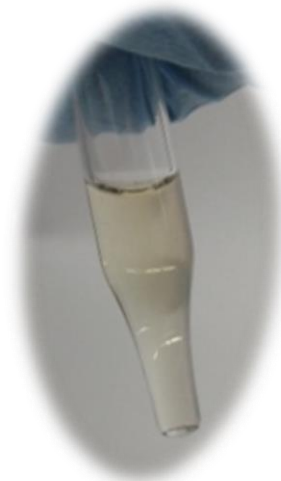
- Seawater ✓
- Estuarine and effluent: dirty extracts
  - Signal suppression ( $> 50\%$ )



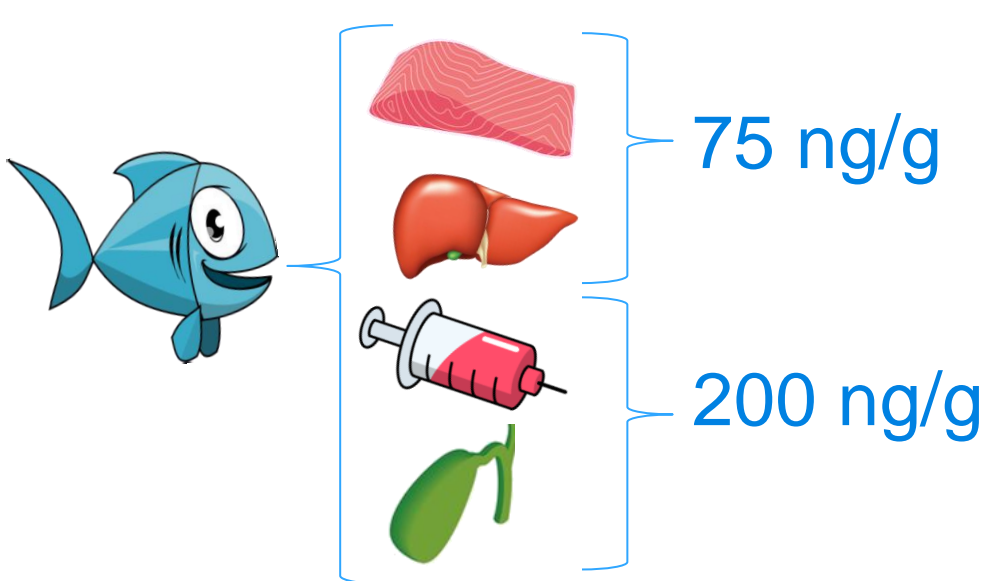
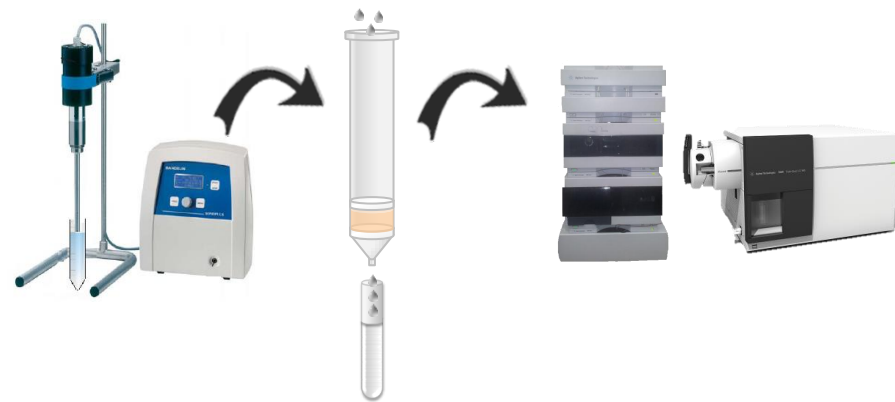
- Solution:  
WAX + HLB



- Cleaner extracts and less signal suppression ( $< 33\%$ )

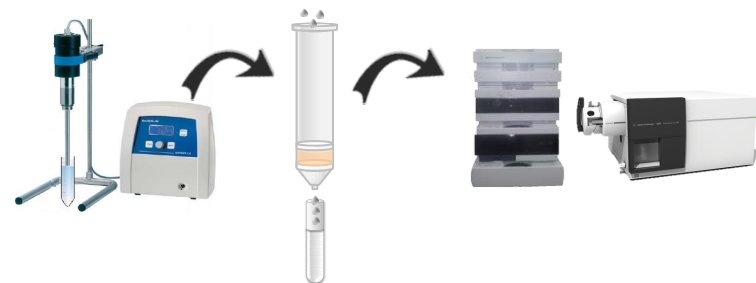


# Method validation





# Method validation

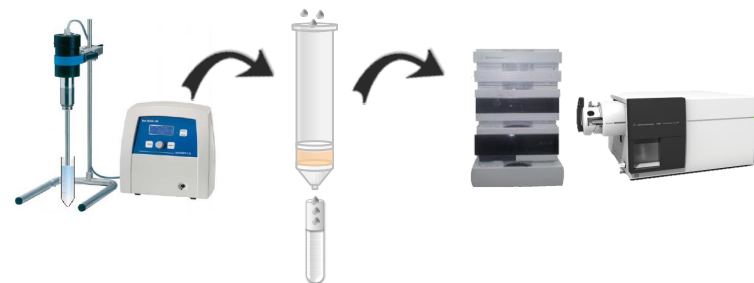


- Linearity
  - $R^2$ : 0.997-0.9997
- Limits of detection
  - $LOD < 2 \text{ ug/mL}$
  - $LOQ < 5 \text{ ug/mL}$
- Precision
  - $RSD < 15 \%$
- Accuracy



# Method validation

• Apparent recoveries: 80 - 126 % ✓

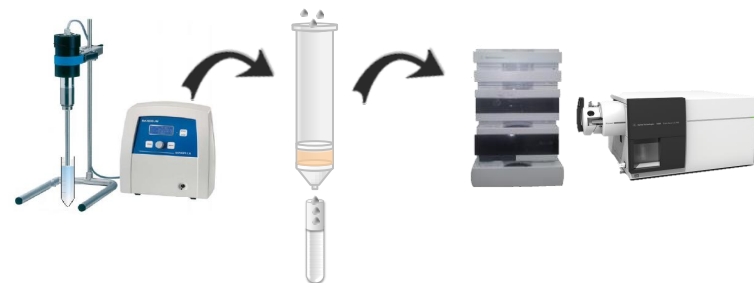


Analyte	Fish tissues		Fish biofluids		Environmental waters		
	Liver	Muscle	Plasma	Bile	Seawater	Estuarine	Effluent
NORF <sup>a</sup>	107	96	93	96	90	84	93
ENO <sup>a</sup>	89	83	95	104	80	82	83
PEFLO <sup>b</sup>	103	89	91	97	101	109	109
OFLO <sup>b</sup>	96	94	110	105	105	93	125
CIPRO <sup>a</sup>	92	92	96	97	87	85	109
DANO <sup>b</sup>	104	92	115	94	112	109	118
LOME <sup>b</sup>	82	94	103	106	106	120	126
ENRO <sup>b</sup>	90	94	100	101	101	97	99
SPAR <sup>a</sup>	14	162	109	70	67	54	59

<sup>a</sup>Corrected with [<sup>2</sup>H<sub>8</sub>]-CIPRO. <sup>b</sup>Corrected with [<sup>2</sup>H<sub>5</sub>]-ENRO.

# Method validation

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DANO <sup>b</sup>	104	92	115	94	112	109	118
LOME <sup>b</sup>	82	94	103	106	106	120	126
ENRO <sup>b</sup>	90	94	100	101	101	97	99
SPAR <sup>a</sup>	14 ✗	162 ✗	109	70	67 ✗	54 ✗	59 ✗

<sup>a</sup>Corrected with [<sup>2</sup>H<sub>8</sub>]-CIPRO. <sup>b</sup>Corrected with [<sup>2</sup>H<sub>5</sub>]-ENRO.

# Results and Discussion

1

Optimization LC-MS/MS analysis of FQs

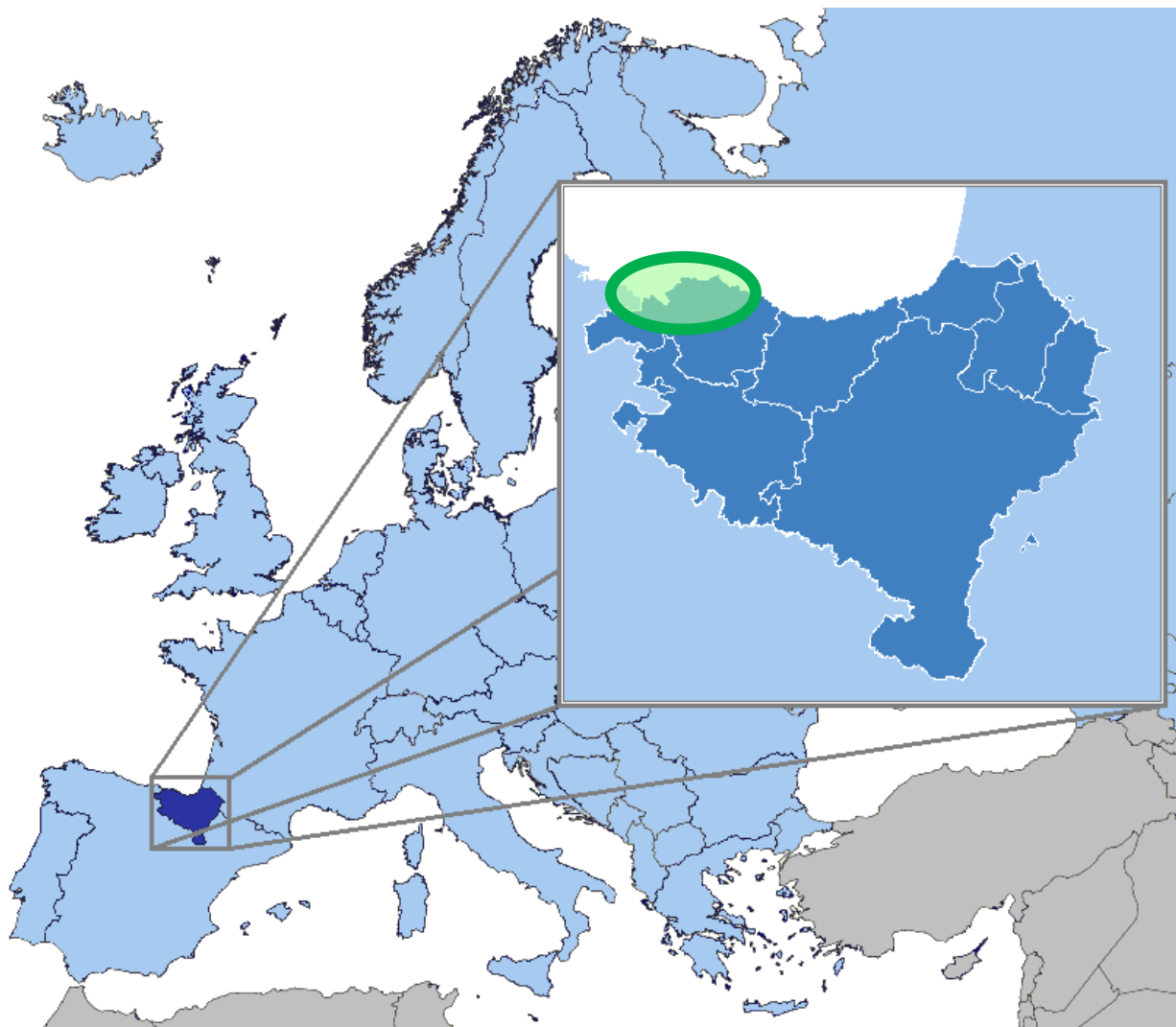
2

Optimization of the extraction: fish tissue/biofluids + environmental water

3

Application to real samples

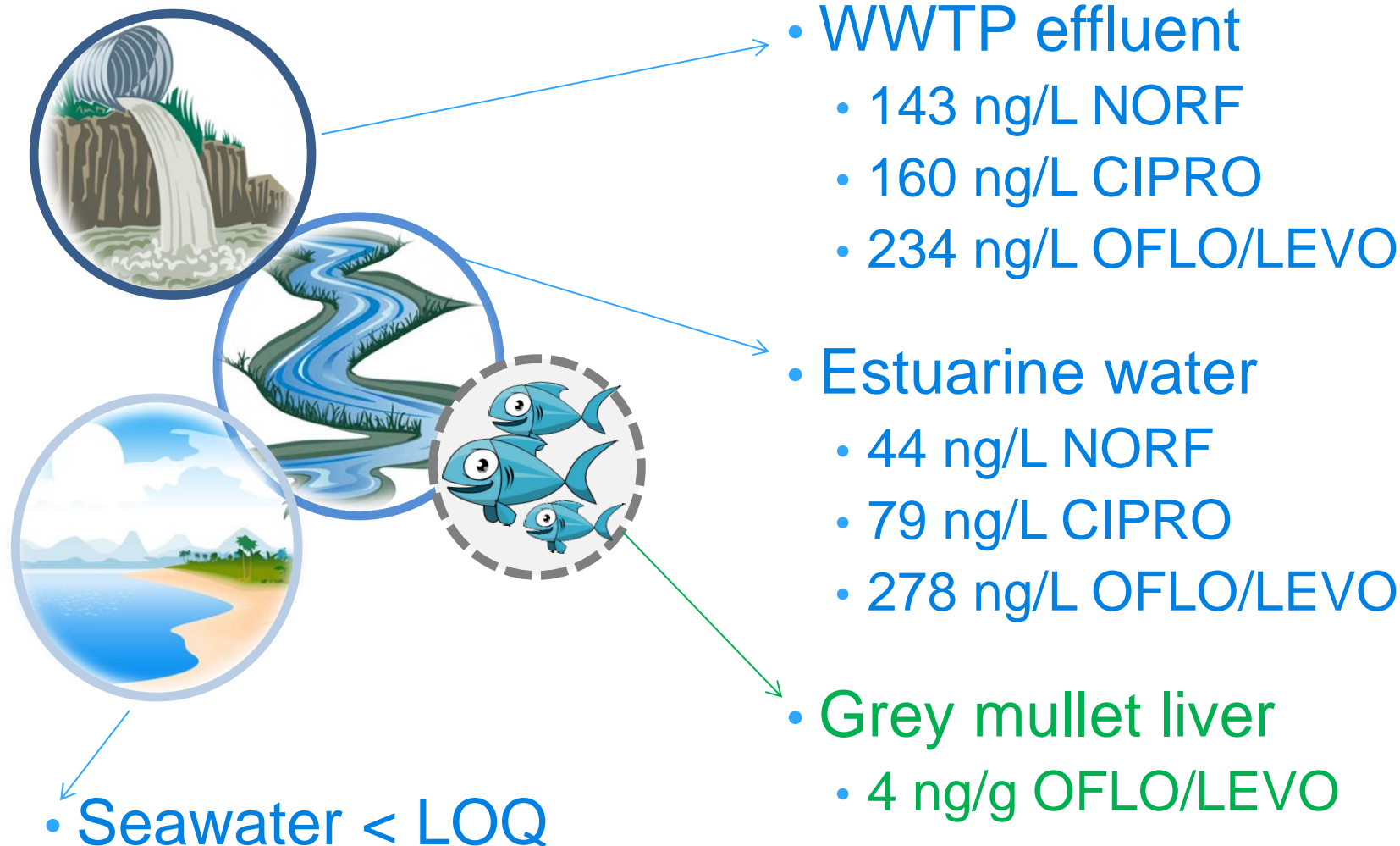
# Real Samples from Biscay Coast



# Real Samples from Biscay Coast



# Real Samples from Biscay Coast

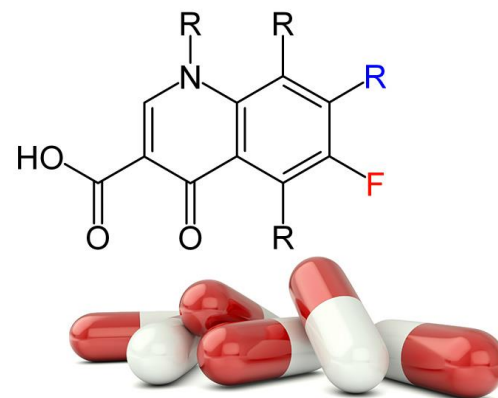


# Conclusions



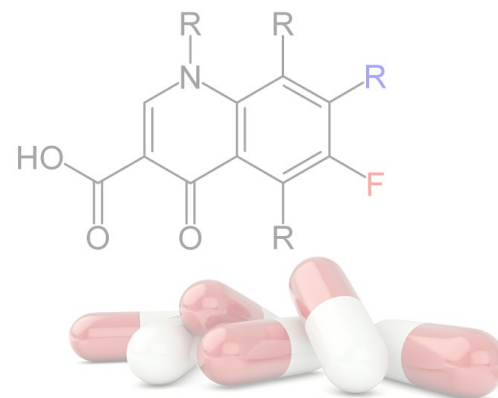
# Conclusions

- Optimization of 10 FQs
  - Correction with labelled analogues



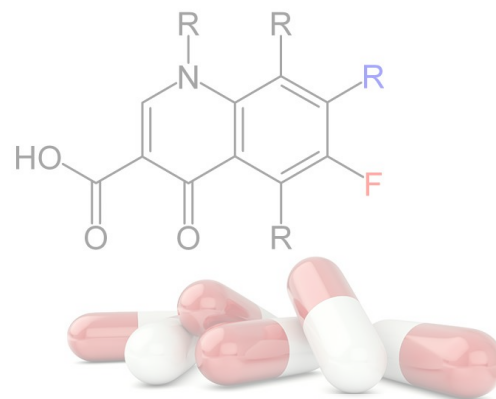
# Conclusions

- Optimization of 10 FQs
  - Correction with labelled analogues
- 7 matrices with different clean-up strategies:
  - Fish muscle and seawater: **RP-SPE**
  - Fish liver: **LLE + RP-SPE**
  - Fish bile and plasma: **MIPs**
  - Estuarine and effluent waters: **WAX + RP-SPE**



# Conclusions

- Optimization of 10 FQs
  - Correction with labelled analogues
- 7 matrices with different clean-up strategies:
  - Fish muscle and seawater: RP-SPE
  - Fish liver: LLE + RP-SPE
  - Fish bile and plasma: MIPs
  - Estuarine and effluent waters: WAX + RP-SPE
- Real samples from the Biscay Coast:
  - CIPRO, NORF and OFLO/LEVO
  - Water and fish liver samples





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